Linear Analogues of Acid- and Ester-Terminated Polyamido Dendrimers: Design, Syntheses, and Physical Properties

George R. Newkome,*[a] Charles N. Moorefield, [a] and Jon D. Epperson[b]

Keywords: Hydrogenation / Amines / Dendrimers / Peptides

The synthesis of linear, unnatural amines and carboxylic acids, based on amide connectivity and possessing identical repeat unit architecture to that of analogous dendrimers, was undertaken to assess and compare their physical characteristics. For the series, unexpected insoluble behavior was observed at low molecular weights in contrast to their branched

counterparts or other known linear dendritic analogs. Molecular modeling suggests a high degree of intra- and intermolecular H-bonding for the series.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2003)

Introduction

The interesting physical properties of dendrimers arise in part from their three-dimensional structures.[1-18] Unlike linear polymers, dendrimers possess a highly branched, "tree-like" architecture with a large number of solvent accessible chain-terminating groups. Following the early stages of dendritic growth, the termini increase in number and become major factors in defining physical properties; in contrast, the end groups of linear polymers remain relatively constant during growth and, thus, play a diminishing role with increasing molecular size. These inherent structural differences lead to many of the observed changes that we see in the chemical and physical properties of dendrimers versus linear polymers. Miller and co-workers^[19–21] have demonstrated that 1,3,5-phenylene-based dendrimers are generally more soluble (ca. 106 times) than their oligop-phenylene linear analogs. Likewise, Fréchet et al.[22,23] noticed a similar correlation for their dendritic poly(aryl ester)s. Other physical properties have been monitored such as: size, [24] viscosity, [25] crystallinity, [25] and thermal properties.^[26] For example, Fréchet et al.^[25] deftly synthesized an exact linear isomer of a 5th-generation polyethereal dendrimer for comparison and showed that the dendrimer's hydrodynamic volume was about 30% smaller than its linear analogue; interestingly, the former was also more reactive toward catalytic hydrogenation than its linear counterpart. More recently, an investigation^[27] on the influence of poly(benzyl ether) dendrons vs. their exact linear analogues

Recent reports of unnatural peptides[27-42] and the reported helical γ-tetra-, hexa-, and octapeptides, [43] along with the structural relationship between the monomers used in our $1\rightarrow 3$ and $1\rightarrow 2$ branched dendritic families, have provided the impetus for the creation of the related gem-dimethyl γ-amino acid derivatives. We herein report a new class of linear analogues corresponding to our well-defined acid- and ester-terminated polyamido dendrimers.[44,45] An exponential growth strategy^[46] was employed to synthesize these monodisperse linear dimers, tetramers, and octamers, using standard peptide [i.e., (DCC and 1-HOBT)^[47] and 2ethoxy-*N*-(ethoxycarbonyl)-1,2-dihydroquinoline (EEDQ)^[48]] coupling conditions.^[49] Molecular modeling studies, as well as ¹H NMR spectroscopic data, of these

polyamides suggest that self-aggregation may contribute to the increased insolubility of the larger analogues.

Results and Discussion

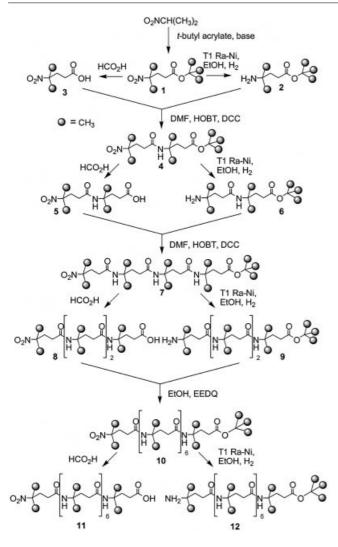
Synthesis of Linear Analogs

The controlled Michael-type addition of tert-butyl acrylate^[50] to 2-nitropropane in refluxing EtOH afforded the desired crystalline nitro ester 1 (Scheme 1), which was supported (13 C NMR) by the single carbonyl peak at $\delta =$ 171.3 ppm and two peaks at $\delta = 87.4 \, (O_2 NC^{4^{\circ}})$ and $\delta =$ 80.7 (Me₃ $C^{4^{\circ}}$) ppm as well as the correct mass (ESI MS) at $m/z = 240.6 \text{ [M + Na]}^+$. Hydrogenation of 1 with T-1 Raney nickel^[51] at 25 °C afforded (95%) the amino ester monomer 2, which was characterized by an upfield shift of the nitro quaternary carbon (13C NMR) resonance from $\delta = 87.4$ ppm to $\delta = 48.7$ ppm (H₂NC^{4°}); no other signifi-

School of Law, George Washington University, Washington, DC 20052, USA

on a porphyrin core was reported; their "results clearly confirm the anticipation that dendrimers are unique when compared to other architectures".

Departments of Chemistry and Polymer Science, The University of Akron, 170 University Cr., Akron, OH 44325-4717, USA Fax: (internat.) + 1-330/972-2535 E-mail: newkome@uakron.edu



Scheme 1. Synthesis of the linear, amide-based unnatural peptides

cant changes were observed indicating that the rest of the molecule was intact. Its 1H NMR signals corresponding to the methyl and methylene groups adjacent to the nitro quaternary carbon atom also shifted upfield from $\delta=1.49$ [O₂NC^{4°}(CH₃)₂] ppm to $\delta=1.21$ [H₂NC^{4°}(CH₃)₂] ppm and from $\delta=2.13$ (O₂NC^{4°}CH₂) ppm to $\delta=1.8$ (H₂NC^{4°}CH₂) ppm, respectively, upon reduction. Care must be taken to keep **2** below 40 $^{\circ}$ C to prevent the facile lactam formation that occurs for γ -amino esters; [52] ester **2** was typically stored at 0 $^{\circ}$ C to minimize cyclization. Treatment of ester **1** with 95% formic acid provided (91%) the monomer **3**, as shown (13 C NMR) by the loss of resonances corresponding to the *tert*-butyl group at $\delta=171.3$ (CO_2R), 80.7 ($C^{4^{\circ}}$ Me₃), and 27.8 [$C^{4^{\circ}}$ (CH_3)₃] ppm and the appearance of a new acid carbonyl peak at $\delta=178.6$ (CO_2H) ppm.

The two key monomers (amine 2 and acid 3) were subjected to amidation by standard peptide coupling conditions (DCC and 1-HOBT)^[47] to generate (40%) dimer 4,

which was evidenced by the notable peak (13 C NMR) shift from $\delta = 48.7$ ($H_2NC^{4^\circ}$) ppm to $\delta = 53.3$ (CONH C^{4°) ppm as well as the correct mass (ESI) value m/z = 331.06 [M + H]⁺.

Catalytic (T-1 Raney nickel) hydrogenation of the nitro moiety in dimer **4** gave rise to the terminal amine **6**, structurally characterized (13 C NMR) by the shift of the signal at $\delta = 87.8$ ($O_2NC^{4^\circ}$) ppm to $\delta = 49.4$ ($H_2NC^{4^\circ}$) ppm, the upfield (1 H NMR) proton shifts at $\delta = 1.49$ [$O_2NC^{4^\circ}(CH_3)_2$] ppm to $\delta = 1.0$ [NH₂C^{4°}(CH₃)₂] ppm, and $\delta = 2.15$ ($O_2NC^{4^\circ}CH_2$) ppm to $\delta = 1.57$ ($H_2NC^{4^\circ}CH_2$) ppm, as well as the correct mass (ESI) at m/z = 301.56 [M + H]⁺. Deprotection of the ester end group of dimer **4** with formic acid provided (90%) the terminal acid **5**, confirmed by the typical loss of the 13 C NMR resonances corresponding to the *tert*-butyl group at $\delta = 170.8$ (CO_2R), 80.6 ($C^{4^\circ}Me_3$), and 28.0 [$C^{4^\circ}(CH_3)_3$] ppm along with the formation of a new signal at $\delta = 178.2$ (CO_2H) ppm.

Combination of amine 6 with acid 5 according to the same coupling conditions (DCC and 1-HOBT) afforded (44%) the desired tetramer 7, as evidenced by the appropriate 13 C NMR shifts caused by amidation: $\delta = 49.4$ $(H_2NC^{4^\circ})$ ppm to $\delta = 53.15$ (CONH C^{4°) ppm. This new tetramer 7 possesses three slightly different CONHC^{4°} peaks at $\delta = 53.2$, 53.15, and 53.1 ppm and exhibits the correct mass (ESI) at $m/z = 557.3 \text{ [M + H]}^+$. Two peaks were also detected at m/z = 501.3 and 454.5 corresponding to the loss of the *tert*-butyl and nitro groups, respectively, which was demonstrated in a related series of building blocks.^[53,54] Catalytic hydrogenation (T-1 Raney nickel) of the nitro terminus of tetramer of 7 generated (95%) the terminal amine 9, whose structural conformation was supported by the shift of the 13 C NMR resonance at $\delta = 87.8$ $(O_2NC^{4^\circ})$ ppm to $\delta = 53.9$ $(H_2NC^{4^\circ})$ ppm, the upfield shift (¹H NMR) for peaks at $\delta = 1.49 [O_2NC^{4\circ}(CH_3)_2]$ ppm to $\delta = 1.15 \, [\text{H}_2 \text{NC}^{4^{\circ}} (\text{C}H_3)_2] \text{ ppm and } \delta = 2.15 \, (\text{O}_2 \text{NC}^{4^{\circ}} \text{C}H_2)$ ppm to $\delta = 1.71 \text{ (H}_2\text{NC}^{4^{\circ}}\text{C}H_2) \text{ ppm, as well as the ob$ served correct mass (ESI) at $m/z = 527.8 \, [M + H]^+$. Deprotection of tetramer 7 with formic acid provided (92%) the acid-terminated tetramer 8, as denoted by the loss (13C NMR) of peaks corresponding to the *tert*-butyl ester at δ = 170.8 (CO_2R), 80.6 (CMe_3), and 28.0 [$C(CH_3)_3$] ppm along with the appearance of a resonance at $\delta = 177.8$ ppm (CO₂H); the amide groups are stable towards these reaction conditions.

Using diverse variations of the DCC and 1-HOBT coupling procedures, reagents **8** and **9** were recovered in toto, in part because these monomers were not readily soluble in DMF. To circumvent this problem, 2-ethoxy-*N*-(ethoxycarbonyl)-1,2-dihydroquinoline (EEDQ) was used to effect the coupling in EtOH affording (74%) the nitro-terminated octamer **10**; no purification was required, since the product cleanly precipitated. This octamer **10** was generally insoluble in most organic solvents but readily dissolved in 95% formic acid, from which the ¹³C and ¹H NMR spectra could be acquired. As expected, acquisition of the NMR spectroscopic data had to be accomplished rapidly to avoid significant *tert*-butyl ester hydrolysis. Pertinent ¹H NMR

peaks for **10** include two broad signals at $\delta = 2.05$ and 1.85 ppm corresponding to the linear chain methylene protons (C⁴°CH₂CH₂), and three distinct methyl proton absorptions at $\delta = 1.28$ [O₂NC(CH₃)₂], 1.14 [C⁴°(CH₃)₃], and 1.04 [HNC⁴°(CH₃)₂] ppm in a 2:3:12 ratio. The ESI mass spectrum of octamer **10** exhibited the correct mass at m/z = 1,009.8 [M + H]⁺.

Catalytic hydrogenation (T-1 Raney nickel) of the nitro terminus in octamer 10 afforded (48%; presumably, increasing chain length might necessitate the use of higher pressures and longer reaction times) the desired terminal amine 12, whose structural conformation was supported by the ¹³C NMR shift of the peak at $\delta = 87.8 \, (O_2 NC^{4^{\circ}})$ ppm to $\delta = 53.9 \, (\mathrm{H_2N}C^{4^\circ})$ ppm and by the observation of the ¹H NMR upfield shifts for the signals at $\delta = 1.28$ $[O_2NC^{4^{\circ}}(CH_3)_2]$ ppm to $\delta = 1.05 [H_2NC^{4^{\circ}}(CH_3)_2]$ ppm and $\delta = 2.05 \text{ (O}_2\text{NC}^{4^{\circ}}\text{C}H_2) \text{ ppm to } \delta = 1.61 \text{ (H}_2\text{NC}^{4^{\circ}}\text{C}H_2)$ ppm. The correct mass (ESI) for the amino ester 12 at $m/z = 980.2 \text{ [M + H]}^+$ was also obtained. Treatment of octamer 10 with formic acid provided (90%) acid-terminated octamer 11, as confirmed by the loss (1H NMR) of the signals corresponding to the tert-butyl group, as noted above, and the correct parent ion (ESI) at m/z = 953.6 [M $+ H1^+$.

In general, these linear analogs of the acid- and esterterminated polyamidoamine dendrimers^[44,45] are less soluble and more crystalline than their dendritic counterparts. This is in good agreement with the previous studies presented by Fréchet^[22] and Miller^[19–21] in aryl, convergently generated dendrimers; however, the physical properties of these linear analogs presented here are quite dramatic. For example, the linear octamer 10, possessing only 8 repeat units (1,009 Da) corresponding to an intermediate size between a first (4 repeats) and second (12 repeats) generation polyamido dendrimer (Figure 1), was insoluble in all nonpolar organic solvents and formed a thermally reversible gel in polar solvents such as DMF and nitrobenzene.

In contrast, the physical properties of the polyethereal linear analogs^[25] do not change significantly until they are as large as a 5th-generation polyethereal dendrimer (31 repeats) with a molecular weight of 6,680 Da. This confirms that the physical behavior of acid- and ester-terminated polyamidoamine dendrimers is quite different from their respective linear analogs. Figure 2 shows elongated ball-andstick models, with exaggerated methyl groups, of the nitro acid tetramer 8 (Figure 2, a) and the nitro acid octamer 11 (Figure 2, b) with measured extended lengths of 27.3 and 53.5 Å, respectively. Figure 2 (c) represents two octamers (i.e., 11; they are shaded differently to aid in visualizing the intertwined chains) minimized together starting from an elongated and perpendicular overlapping conformation. Notably, much of the exterior is occupied by the lipophilic methyl groups; whereas, the heteroatom connectivity is oriented inward. This suggests that the unexpected insolubility at even these low molecular weights might be due to extensive intra- and intermolecular H-bonding. Other data supporting this conclusion includes the downfield shift and broadening of the amide proton absorption ($\delta = 5.85$ ppm

Figure 1. Idealized representations of 1st- and 2nd-generation polypeptide dendrimers for comparison to the linear analogs

to $\delta = 6.05$ ppm) for amino ester **6** when progressing from a dilute to a concentrated sample; this fits with literature precedence.^[55]

The diminished solubility of the octamer 12 prevented its addition to the acid-terminated dendrimers; however, the tetramer 9 was covalently linked to members of the acidterminated dendrimers as demonstrated by the coupling of the known tetraacid chloride^[56] 13 with a slight excess of amine 9 in THF in the presence of Et₃N to afford (62%) the 1st-generation dendritic core 14 (Scheme 2). The presence (13 C NMR) of two carbonyl peaks at $\delta = 173.2$ (CONH) and 170.9 ppm (CO_2R); three quaternary carbon peaks at $\delta = 80.1$ (Me₃C), 52.8 (four overlapped HNC^{4°}), and 45.2 ppm ($C^{4\circ}CH_2$); seven distinct methylene peaks at $\delta = 68.9 (C^{4^{\circ}}CH_2O), 67.4 (OCH_2CH_2), 37.0 (OCH_2CH_2),$ 36.1 and 34.7 (C⁴°CH₂CH₂), 31.8 and 30.5 (C⁴°CH₂CH₂) ppm; and three methyl peaks at $\delta = 27.9 \, [C^{4^{\circ}}(CH_3)_3], 26.5$ and 26.4 [C^{4°}(CH₃)₂] ppm as well as ESI MS possessing m/ z = 1230.7 corresponding to the doubly charged parent ion

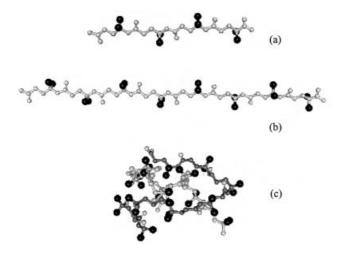
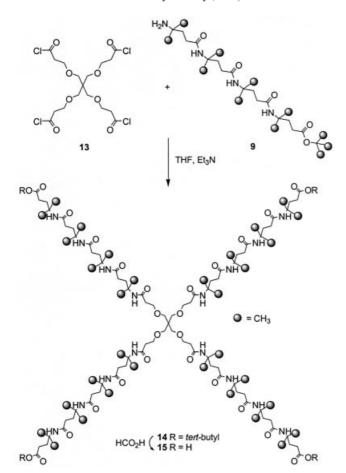


Figure 2. Ball-and-stick representations of the nitro acids 8 (a) and 11 (b) with measured lengths of 27.3 and 53.5 Å, respectively, and two octamers 11 (c) minimized together resulting in aggregation (modeling, minimization, and dynamics were performed with Cerius2 version 4.6 software by Accelrys, Inc.)



Scheme 2. Facile attachment of the tetramer 9 to a poly(acid chloride)

[M + 2 H]²⁺ supported the transformation. Ester **14** was subsequently hydrolyzed with formic acid to give (92%) the tetraacid **15**, as suggested by the loss of the signature absorptions corresponding to the *tert*-butyl groups along with the formation of a new acid peak at $\delta = 177.5$ ppm

(CO₂H). The tetraacid **15** also showed (ESI) the parent ion peak at $m/z = 1117.7 \text{ } [\text{M} + 2 \text{ H}]^{2+}.$

Conclusion

These simple, linear polyamides are but new members of a growing family of extended, unnatural polypeptides, which are physically interesting as related to either their highly branched or natural, well-known counterparts. Even at low molecular weights and low repeat unit number, these materials show unexpectedly poor solubility in nonpolar solvents due to H-bonding interactions, an attribute of structural self-assembly. The complementary amine and ester functionalities make these materials ideal for use as spacer moieties in dendritic constructs.

Experimental Section

General Remarks: Melting point data were obtained in capillary tubes with an Electrothermal 9100 melting point apparatus and are uncorrected. ¹H and ¹³C NMR spectra were obtained in CDCl₃, except where noted, and are recorded at 250.13 MHz. The ¹H and ¹³C NMR spectra of samples dissolved in formic acid were obtained without a lock signal and were shimmed on the FID. Infrared spectra (IR) were obtained (KBr pellet, unless otherwise noted) and recorded with an ATI Mattson Genesis Series FTIR spectrometer. Mass spectrometric data were obtained using an Esquire electron ionization mass spectrometer (ESI) and are reported as: (relative intensity) [assignment]. ESI samples were typically prepared in MeOH/H₂O/TFA (70:30:01) for positive ion mode or Me₂₋ CHOH/H₂O/NH₃ (70:30:1) for negative ion mode. The 8-mer nitro/ ester, nitro/acid, and amino/ester samples were all first dissolved in 95% formic acid and then diluted 1:10 in MeOH/H2O (4:1) before loading into the ESI.

tert-Butyl 4,4-Dimethyl-4-nitropentanoate (1): A total of 1.2 equiv. (0.2 equiv. excess) of *tert*-butyl acrylate (172.6 g, 1.35 mol) was added portionwise over 10 min to a refluxing solution of 2-nitropropane (100 g, 1.12 mol) and Triton B (2.2 mL, 40% in MeOH) in refluxing abs. EtOH (500 mL). After the *tert*-butyl acrylate was added, additional Triton B (2.2 mL) was added and the mixture was refluxed for 14 h. The solvent was removed in vacuo to afford a crude light-yellow solid, which was recrystallized with EtOH/H₂O to give (60%) the desired ester: 146 g; m.p. 63.7–65.1 °C. ¹³C NMR: δ = 171.3 (C=O), 87.4 (O₂NC^{4°}), 80.7 (CMe₃), 35.3 (C^{4°}CH₂), 30.3 (C^{4°}CH₂CH₂), 27.8 [C(CH₃)₃], 25.6 [C^{4°}(CH₃)₂], ¹H NMR: δ = 2.13 (br., 4 H, CH₂CH₂), 1.49 [s, 6 H, C^{4°}(CH₃)₂], 1.34 [s, 9 H, C^{4°}(CH₃)₃] ppm. IR (KBr): \tilde{v} = 1730 (ester C=O), 1540 (NO₂), 1373 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 240.6 (100) [M + Na]⁺.

tert-Butyl 4,4-Dimethyl-4-aminopentanoate (2): A solution of nitro ester 1 (40 g, 184 mmol) in abs. EtOH (300 mL) with T-1 Raney Ni (30 g, 50:50% Al/Ni) was hydrogenated (3.79 × 10⁵ Pa) at 25 °C for 12 h. The solution was filtered through Celite to remove the catalyst, using care to not let the filter pad become dry due to the pyrophoric nature of the catalyst, and the solvent was removed in vacuo to give (95%) the amino ester 2, as a clear liquid: 32.7 g. 13 C NMR: δ = 173.1 (C=O), 79.6 (CMe₃), 48.7 (H₂NC^{4°}), 39.1 (C^{4°}CH₂), 30.8 (C^{4°}CH₂CH₂), 30.0 [C^{4°}(CH₃)₂], 27.7 [C(CH₃)₃]. 14 H NMR: δ = 2.35 (t, J = 8.3 Hz, 2 H, $^{4°}$ CH₂CH₂), 1.8 (t, J = 8.3 Hz, 2 H, $^{4°}$ CH₂CH₂), 1.55 [s, 9 H, $^{4°}$ C(CH₃)₃], 1.44 (s, 2 H,

N*H*₂), 1.21 [s, 6 H, C^{4°}(C*H*₃)₂] ppm. IR (neat): \tilde{v} = 3355 (NH₂), 3287 (NH₂), 1729 (ester C=O) cm⁻¹. MS (ESI): m/z (%) = 188.2 (41.2) [M + H]⁺, 132.3 (100) [M + H - CMe₃]⁺.

4,4-Dimethyl-4-nitropentanoic Acid (3): Ester **1** (40 g, 184 mmol) was dissolved in formic acid (95%, 200 mL) and stirred at 25 °C for 12 h. The excess formic acid was removed in vacuo, water was added (200 mL), and the solution was again concentrated in vacuo to ensure the absence of any residual formic acid affording (91%) the acid **3**, as a white solid: 27 g; m.p. 150–151 °C. ¹³C NMR: δ = 178.6 (CO_2H), 87.2 (O_2NC^4 °), 34.8 (C^4 °CH₂), 29.1 (C^4 °CH₂CH₂), 25.7 [C^4 °(CH_3)₂] ppm. ¹H NMR: δ = 11.2 (br., 1 H, CO_2H), 2.29 (m, 2 H, C^4 °CH₂), 2.16 (m, 2 H, C^4 °CH₂CH₂), 1.47 [s, 6 H, C^4 °(CH_3)₂] ppm. IR (KBr): $\tilde{v} = 3500-3000$ (CO_2H), 1674 (C=O), 1563 (NO_2), 1367 (NO_2) cm⁻¹. MS (ESI): m/z (%) = 160.8 (100) [M - H]⁺.

2-mer 4: A stirred mixture of nitro acid 3 (10.95 g, 68 mmol), 1-HOBT (9.18 g, 68 mmol), and DCC (14.0 g, 68 mmol) in DMF (100 mL) was maintained at 25 °C for 30 min, before amine 2 (12.71 g, 68 mmol) was added. The mixture was stirred for 18 h, then filtered through Celite to remove the dicyclohexylurea. The solvent was removed in vacuo to give the crude product, which was dissolved in CH₂Cl₂ (100 mL) and washed with water (3 ×, 200 mL). After concentration in vacuo, the residue was recrystallized (cyclohexane) to give (40%) the desired amide 4, as colorless crystals: 8.9 g; m.p. 83-84.5 °C. 13 C NMR: $\delta = 173.5$ (CONH), 170.8 (CO₂R), 87.8 (O₂N $C^{4\circ}$), 80.6 (CMe₃), 53.3 $(O_2NC^{4^{\circ}}CH_2)$, 35.1 (HNC^{4°}CH₂), 31.8 $(HNC^{4^{\circ}}), 35.9$ $(CH_2CONH),$ 30.6 (CH_2CO_2R) , 28.0 $[C(CH_3)_3]$, 26.5 $[NHC^{4^{\circ}}(\mathit{CH}_{3})_{2}],\ 25.7\ [O_{2}NC^{4^{\circ}}(\mathit{CH}_{3})_{2}]\ ppm.\ ^{1}H\ NMR:\ \delta\ =\ 5.84$ (NH), 2.15 (m, 4 H, O₂NC⁴°CH₂, CH₂CO₂R), 2.01 (m, 2 H, $HNC^{4^{\circ}}CH_2$), 1.84 (t, J = 7.4 Hz, 2 H, $HNC^{4^{\circ}}CH_2$), 1.49 [s, 6 H, $O_2NC^{4^{\circ}}(CH_3)_2$, 1.35 [s, 9 H, $C^{4^{\circ}}(CH_3)_3$], 1.22 [s, 6 H, $HNC^{4^{\circ}}(CH_3)_2$ ppm. IR (KBr): $\tilde{v} = 3333$ (NH), 1689, 1667 (ester/ amide C=O), 1538 (NO₂), 1366 (NO₂) cm⁻¹. MS (ESI): m/z (%) = $353.19 (10.3) [M + Na]^+, 275.13 (100) [M + H - CMe_3]^+.$

2-mer Acid 5: Amide **4** (4 g, 12 mmol) was dissolved in formic acid (95%, 200 mL) and stirred at 25 °C for 12 h. The workup was as for **3** above to give (90%) **5**, as a pure white solid: 3.0 g; m.p. 120-122 °C. 13 C NMR: δ = 178.2 (CO₂H), 171.7 (CONH), 87.9 (O₂NC⁴°), 53.4 (HNC⁴°), 36.0, 34.7 (C⁴°CH₂), 31.7, 29.2 (C⁴°CH₂CH₂), 26.5, 25.6 [C⁴°(CH₃)₂] ppm. 1 H NMR: δ = 11.0 (br., 1 H, CO₂H), 6.15 (br., 1 H, CONH) 2.21 (t, J = 7.2 Hz, 2 H, CH₂CO₂), 2.05 (m, 2 H, O₂NC⁴°CH₂), 1.95 (m, 2 H, CH₂NH), 1.88 (t, J = 7.4 Hz, 2 H, HNC⁴°CH₂), 1.43 [s, 6 H, O₂NC⁴°(CH₃)₂], 1.17 [s, 6 H, HNC⁴°(CH₃)₂] ppm. IR (KBr): $\tilde{v} = 3384$ (NH), 3200-2900 (CO₂H), 1738 (acid C=O), 1538 (NO₂) cm⁻¹. MS (ESI): mlz (%) = 297.1 (55.7) [M + Na]⁺, 275.2 (100) [M + H]⁺.

2-mer Amine 6: A solution of the ester **4** (4 g, 12 mmol) in abs. EtOH (300 mL) with T-1 Raney Ni [10 g, Al/Ni (1:1)] was hydrogenated (3.79 × 10⁵ Pa) at 25 °C for 12 h. The solution was carefully filtered, as in the procedure for compound **2**, through Celite, then concentrated in vacuo to give (92%) the amino ester **6**: 3.4 g; m.p. 68.5–70 °C. ¹³C NMR: δ = 173.6 (CONH), 172.9 (CO₂R), 80.6 (CMe₃), 53.1 (CONHC^{4°}), 49.4 (H₂NC^{4°}), 39.9 (H₂NC^{4°}CH₂), 35.2 (NHC^{4°}CH₂CH₂), 33.0 (CH₂CONH), 30.7 (CH₂CO₂R), 30.4 [H₂NC^{4°}(CH₃)₂], 28.1 [C(CH₃)₃], 26.7 [NHC^{4°}(CH₃)₂] ppm. ¹H NMR: δ = 5.88 (CON*H*), 2.1 (m, 4 H, C^{4°}CH₂CH₂), 1.84 (t, *J* = 7.8 Hz, 2 H, HNC^{4°}CH₂), 1.31 [s, 9 H, C^{4°}(CH₃)₃], 1.21 [s, 6 H, HNC^{4°}(CH₃)₂], 1.0 [s, 6 H, H₂NC^{4°}(CH₃)₂] ppm. IR (neat): \tilde{v} = 3313 (NH₂), 1726 (ester C=O), 1646 (amide C=O) cm⁻¹. MS

(ESI): m/z (%) = 323.28 (6.1) [M + Na]⁺, 301.56 (100) [M + H]⁺.

4-mer 7: A stirred mixture of acid 5 (6.35 g, 23 mmol), 1-HOBT (3.13 g, 23 mmol), and DCC (4.8 g, 23 mmol) in DMF (100 mL) was maintained at 25 °C for 30 min, then amine 6 (6.95 g, 23 mmol) was added. The mixture was stirred for an additional 18 h. The solution was subsequently filtered through Celite and concentrated in vacuo to give the crude product, which was dissolved in CH₂Cl₂ (100 mL) and washed with 5% HCl (3×200 mL), then aq. NaOH (5%, 200 mL). The solvent was removed in vacuo to afford the tetramer 7, which was recrystallized (EtOAc) to give (44%) the desired product: 5.6 g; m.p. 189.3–190.3 °C. ¹³C NMR: $\delta = 173.5$, 173.4, 173.3 (CONH), 170.8 (CO_2R), 87.8 ($O_2NC^{4^\circ}$), 80.6 (CMe_3), 53.2, 53.15, 53.1 (HNC 4 °), 35.9, 35.1 (C 4 °CH $_2$), 31.8, 30.6 $(C^{4^{\circ}}CH_2CH_2)$, 28.0 $[C(CH_3)_3]$, 26.5, 25.7 $[C^{4^{\circ}}(CH_3)_2]$ ppm. ¹H NMR: $\delta = 7.00$, 6.92 (br., NH), 2.14 (m, 8 H, CH₂CH₂), 2.05 (m, 2 H, $O_2NC^{4\circ}CH_2$), 1.85–1.75 (m, J = 7.4 Hz, 6 H, $HNC^{4\circ}CH_2$) 1.48 [s, 6 H, $O_2NC^{4^{\circ}}(CH_3)_2$], 1.35 [s, 9 H, $C(CH_3)_3$], 1.22 [br, 18 H, $HNC^{4^{\circ}}(CH_3)_2$] ppm. IR (KBr): $\tilde{v} = 3284$ (NH), 1729 (ester C=O), 1642 (amide C=O), 1546 (NO₂), 1369 (NO₂) cm⁻¹. MS (ESI): m/ z (%) = 557.3 (45.6) [M + H]⁺, 501.3 (100) [M + H - CMe₃]⁺.

4-mer Acid 8: Tetramer **7** (5 g, 90 mmol) was dissolved in formic acid (95%, 200 mL) and stirred at 25 °C for 14 h. The workup was similar to that of **3**, and afforded (92%) acid **8**, as a pure white solid: 4.1 g; m.p. 176.3–177.7 °C. ¹³C NMR (CD₃OD): δ = 177.8 (CO₂H), 175.8, 173.8 (CONH), 89.0 (O₂NC^{4°}), 54.3, 54.2, 54.0 (HNC^{4°}), 37.5, 37.3, 37.2, 35.7 (C^{4°}CH₂), 33.1, 33.0, 32.6, 30.5 (C^{4°}CH₂CH₂), 27.2, 27.1, 26.1 [C^{4°}(CH₃)₂] ppm. ¹H NMR (CD₃OD): δ = 11.0 (br., 1 H, CO₂H), 6.15 (br., 3 H, CONH) 2.2 (m, 12 H), 1.9 (m, 4 H), 1.58 [s, 6 H, O₂NC^{4°}(CH₃)₂], 1.28 [s, 18 H, HNC^{4°}(CH₃)₂] ppm. IR (KBr): \tilde{v} = 3500–3300 (CO₂H), 3279 (NH), 1711 (acid C=O), 1642 (amide C=O), 1544 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 501.2 (100) [M + H]⁺.

4-mer Amine 9: A solution of nitro ester **7** (5 g, 90 mmol) in abs. EtOH (300 mL) with T-1 Raney Ni [10 g, Al/Ni (1:1)] was hydrogenated (3.79 × 10⁵ Pa) at 25 °C for 14 h. The solution was filtered, as in the procedure for compound **2**, through Celite and the solvent was removed in vacuo to give (95%) the amino ester **9**, as a white solid: 4.5 g; m.p. 209.7–210.7 °C. ¹³C NMR (CD₃OD): δ = 175.8, 175.7, 174.9 (CONH), 81.4 (CMe₃), 54.1, 53.9 (NHC^{4°}), 41.4, 37.3, 37.2, 35.7 (C^{4°}CH₂), 33.5, 33.1, 31.8 (C^{4°}CH₂CH₂), 29.8, 28.7, 28.5, 27.3 [C^{4°}(CH₃)₂], 27.1 [C(CH₃)₃] ppm. ¹H NMR (CD₃OD): δ = 7.6, 7.5, 7.4 (CON*H*), 2.15 (m, 8 H, C*H*₂CO), 1.95 (m, 6 H, HNC^{4°}C*H*₂), 1.71 (m, 2 H, H₂NC^{4°}C*H*₂), 1.48 [s, 9 H, C(C*H*₃)₃], 1.33, 1.32 [s, 18 H, HNC^{4°}(C*H*₃)₂], 1.15 [s, 6 H, H₂NC^{4°}(C*H*₃)₂] ppm. IR (KBr): \tilde{v} = 3274 (NH), 1726 (ester C=O), 1642 (amide C=O), 1566 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 527.8 (100) [M + H]⁺.

8-mer 10: Nitro acid **8** (100 mg, 2 mmol) and amine **9** (105 mg, 2 mmol) were dissolved in EtOH (ca. 5 mL) and refluxed, then cooled and 2-ethoxy-*N*-(ethoxycarbonyl)-1,2-dihydroquinoline (EEDQ; 54 mg, 22 mmol) was added; the mixture was stirred at 50 °C for 14 h. The mixture was subsequently cooled to 25 °C and the resultant white precipitate was filtered and washed with EtOH (3 × 5 mL) to yield (74%) the desired 8-mer **10**: 150 mg; m.p. 250–251.5 °C. ¹³C NMR (HCO₂H): δ = 176.9 (CONH), 173.6 (CO₂R), 87.8 (O₂NC^{4°}), 82.5 (CMe₃), 54.6 (HNC^{4°}), 35.8, 35.0 (C^{4°}CH₂), 31.2, 30.3 (C^{4°}CH₂CH₂), 26.8 [C(CH₃)₃], 25.4, 24.4, 19.7 [C^{4°}(CH₃)₂] ppm. ¹H NMR (HCO₂H): δ = 7.18, 7.16, 7.1, 6.95 (br., 7 H, N*H*), 2.05, 1.85 (br., 32 H, C*H*₂C*H*₂), 1.28 [s, 6 H, O₂NC^{4°}(C*H*₃)₂], 1.14 [s, 9 H, C(C*H*₃)₃], 1.04 [s, 42 H, HNC^{4°}(C*H*₃)₂] ppm. IR (KBr): \tilde{v} = 3280 (NH), 1730 (ester C=O),

1641 (amide C=O), 1552 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 1047.7 (100) [M + K]⁺, 1009.8 (34.1) [M + H]⁺.

8-mer Acid 11: A stirred mixture of ester 8 (120 mg, 12 mmol) in formic acid (95%, 10 mL) was maintained at 25 °C for 14 h, then worked up as for 3 affording (90%) the desired acid 11, as a pure white solid: 100 mg; m.p. 246–248 °C. 13 C NMR (HCO₂H): δ = 179.1 (CO₂H), 176.9 (CONH), 87.8 (O₂NC⁴°), 54.6, 54.3 (HNC⁴°), 35.8, 35.0, 33.3 (C⁴°CH₂), 31.1, 28.6 (C⁴°CH₂CH₂), 25.4, 24.4, 19.7 [C⁴°(CH₃)₂] ppm. 1 H NMR (HCO₂H): δ = 7.18, 7.20, 6.95 (br., 7 H, N*H*), 2.07, 1.85 (br., 32 H, C*H*₂C*H*₂), 1.28 [s, 6 H, O₂NC⁴°(C*H*₃)₂], 1.04 [s, 42 H, HNC⁴°(C*H*₃)₂] ppm. IR (KBr): \tilde{v} = 3500–3300 (CO₂H), 3280 (NH), 1717 (acid C=O), 1641 (amide C=O), 1552 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 975.4 (5.3) [M + Na]⁺, 953.6 (100) [M + H]⁺.

8-mer Amine 12: A stirred solution of ester 10 (150 mg, 15 mmol) in abs. EtOH (100 mL) with T-1 Raney Ni [2 g, Al/Ni (50:50)] was hydrogenated (1.38 \times 10⁶ Pa) at 115 °C for 14 h in a metal bomb. The solution was filtered, as in the procedure for compound 2, through Celite and the solvent was removed in vacuo to give (48%) the amino ester 12, as a white solid: 70 mg; m.p. 245-246.5 °C. ¹³C NMR (HCO₂H): $\delta = (CONH)$, 81.4 ($C^{4\circ}Me_3$), 54.1, 53.9 $(HNC^{4^{\circ}})$, 41.4, 37.3, 37.2, 35.7 $(C^{4^{\circ}}CH_2)$, 33.5, 33.1, 31.8 $(C^{4^{\circ}}CH_2CH_2)$, 29.8, 28.7, 28.5, 27.3 $[C^{4^{\circ}}(CH_3)_2]$, 27.1 $[C(CH_3)_3]$ ppm. ¹H NMR (HCO₂H): $\delta = 7.5, 7.4, 7.3$ (CONH), 2.05 (m, 8 H, CH_2CO), 1.85 (m, 6 H, $HNC^{4}{}^{\circ}CH_2$), 1.61 (m, 2 H, $H_2NC^{4^{\circ}}CH_2$), 1.38 [s, 9 H, $C(CH_3)_3$], 1.23, 1.22 [s, 18 H, $HNC^{4^{\circ}}(CH_3)_2$], 1.05 [s, 6 H, $H_2NC^{4^{\circ}}(CH_3)_2$] ppm. IR (KBr): $\tilde{v} =$ 3279 (NH), 1731 (ester C=O), 1641 (amide C=O), 1553 (NO₂) cm⁻¹. MS (ESI): m/z (%) = 1008.0 (77.0) [M + Na]⁺, 980.2 (100) $[M + H]^{+}$.

- 4 × 4 Ester 14: To a refluxing solution of amine 9 (1.53 g, 2.9 mmol) and Et₃N (300 mg, 2.9 mmol) in dry THF (50 mL) was added dropwise, the tetraacid chloride 13 (320 mg, 646 mol), prepared from the corresponding tetracarboxylic acid,[57] in THF (10 mL) over 30 min. The solution was then refluxed for 14 h then the solvent was removed in vacuo to give the crude product, which was dissolved in MeOH (5 mL) and dialyzed against MeOH (3 \times 1000 mL) using a 1000 MWCO dialysis membrane (regenerated cellulose) to afford (62%) the ester 14: 980 mg; m.p. 177-179 °C. ¹³C NMR: $\delta = 173.2$ (CONH), 170.9 (CO₂R), 80.1 (CMe₃), 68.9 $(C^{4^{\circ}}CH_{2}O)$, 67.4 $(OCH_{2}CH_{2})$, 52.8 $(HNC^{4^{\circ}})$, 45.2 $(C^{4^{\circ}}CH_{2})$, 37.0 (OCH_2CH_2) , 36.1, 34.7 $(C^{4^{\circ}}CH_2CH_2)$, 31.8, 30.5 $(C^{4^{\circ}}CH_2CH_2)$, 27.9 [C(CH₃)₃], 26.5, 26.4 [C^{4°}(CH₃)₂] ppm. ¹H NMR: $\delta = 7.0$, 6.97, 6.32 (br., 12 H, CONH), 3.52 (br., 8 H, OCH₂CH₂), 3.22 (br., 8 H, $C^{4\circ}CH_2O$), 2.23 (br., 8 H, OCH_2CH_2), 2.1 (m, 32 H, $C^{4\circ}CH_2CH_2$), 1.85 (m, 32 H, $C^{4\circ}CH_2CH_2$), 1.32 [s, 36 H, $C^{4^{\circ}}(CH_3)_3$, 1.20 [s, 96 H, $C^{4^{\circ}}(CH_3)_2$] ppm. IR (KBr): $\tilde{v} = 3307$ (NH), 1732 (ester C=O), 1647 (amide C=O), 1547 cm⁻¹. MS (ESI): m/z (%) = 1230.7 (100) [M + 2 H]²⁺.
- **4 × 4 Acid 15:** A stirred mixture of ester **14** (880 mg, 36 mmol) in formic acid (95%, 10 mL) was maintained at 25 °C for 15 h, then worked up as described above for **3**, to yield (92%) the desired acid **15**, as a pure white solid: 730 mg; m.p. 159–161 °C. ¹³C NMR (CD₃OD): δ = 177.5 (CO₂H), 175.6, 173.4 (CONH), 70.6 (C⁴°CH₂O), 68.9 (OCH₂), 54.2, 54.1 (HNC⁴°), 46.6 (C⁴°CH₂), 38.5 (OCH₂CH₂), 37.2, 35.6 (C⁴°CH₂), 33.1, 30.4 (C⁴°CH₂CH₂), 27.2, 27.1 [C⁴°(CH₃)₂]. ¹H NMR (CD₃OD): δ = 7.58 (br., 16 H, CON*H*), 3.65 (br., 8 H, OCH₂CH₂), 3.4 (br., 8 H, C⁴°CH₂O), 2.4 (br., 8 H, OCH₂CH₂), 2.2 (br., 32 H, C⁴°CH₂CH₂), 2.05 (br., 32 H, C⁴°CH₂CH₂), 1.30 [s, 96 H, C⁴°(CH₃)₂]. IR (KBr): \tilde{v} = 3315 (NH), 3082, 1715 (acid C=O), 1651 (amide C=O) cm⁻¹. MS (ESI): *m/z* (%) = 1117.7 (100) [M + 2 H]²⁺.

Acknowledgments

We gratefully thank the National Science Foundation (DMR-9901393 & 0196231), the Office of Naval Research (N00013-99-1-0082), and the Ohio Board of Regents for financial support.

- [1] G. R. Newkome, C. N. Moorefield, F. Vögtle, Dendrimers and Dendrons: Concepts, Syntheses, Applications, Wiley-VCH, Weinheim, Germany 2001.
- [2] Dendrimers and Other Dendritic Polymers (Eds.: J. M. J. Fréchet, D. A. Tomalia), John Wiley & Sons, West Sussex, UK, 2001.
- [3] A. W. Bosman, H. M. Janssen, E. W. Meijer, Chem. Rev. 1999, 99, 1665-1688.
- [4] I. P. Beletskaya, A. V. Chuchurjukin, Russ. Chem. Rev. 2000, 69, 639-660.
- [5] A.-M. Caminade, R. Laurent, B. Chaudret, J.-P. Majoral, Coord. Chem. Rev. 1998, 178–180, 793–821.
- [6] J. M. J. Fréchet, C. J. Hawker, in *Comprehensive Polymer Chemistry*, 2nd suppl. (Eds.: S. L. Aggarwal, S. Russo), Elsevier, Oxford, UK, 1996, pp. 71–132.
- [7] C. J. Hawker, W. Devonport, in Step-Growth Polymers for High-Performance Materials New Synthetic Methods (Eds.: J. L. Hedrick, J. W. Labadie), American Chemical Society, Washington, D. C., 1996, pp. 186–196.
- [8] B. Klajnert, M. Bryszewska, Acta Biochim. Pol. 2001, 48, 199-208.
- [9] L. J. Twyman, A. S. H. King, I. K. Martin, Chem. Soc. Rev. 2002, 31, 69–82.
- [10] M. Seiler, Chem. Eng. Technol. 2002, 25, 237-253.
- [11] I. Gitsov, in Advances in Dendritic Macromolecules (Ed.: G. R. Newkome), Elsevier Science Ltd., Kidlington, Oxford, UK, 2002, pp. 45–87.
- [12] S. M. Grayson, J. M. J. Fréchet, Chem. Rev. 2001, 101, 3819-3867.
- [13] D. Astruc, F. Chardac, Chem. Rev. 2001, 101, 2991-3023.
- [14] A. M. Muzafarov, E. A. Rebrov, Vysokomol. Soedin., Ser. A 2000, 42, 2015–2040.
- [15] K. Inoue, Prog. Polym. Sci. 2000, 25, 453-571.
- [16] F. Vögtle, S. Gestermann, R. Hesse, H. Schwierz, B. Windisch, Prog. Polym. Sci. 2000, 25, 987-1041.
- [17] S. Hecht, J. M. J. Fréchet, Angew. Chem. Int. Ed. 2001, 40, 75-91.
- ^[18] M. W. P. L. Baars, E. W. Meijer, *Top. Curr. Chem.* **2000**, *210*, 131–227
- [19] T. M. Miller, T. X. Neenan, *Chem. Mater.* **1990**, *2*, 346–349.
- [20] T. M. Miller, T. X. Neenan, R. Zayas, H. E. Bair, J. Am. Chem. Soc. 1992, 114, 1018-1025.
- [21] T. M. Miller, E. W. Kwock, T. X. Neenan, *Macromolecules* 1992, 25, 3143-3148.
- [22] K. L. Wooley, J. M. J. Fréchet, C. J. Hawker, *Polymer* 1994, 35, 4489–4495.
- [23] C. J. Hawker, J. M. J. Fréchet, in Step-Growth Polymers for High-Performance Materials New Synthetic Methods (Eds.: J. L. Hedrick, J. W. Labadie), American Chemical Society, Washington, D. C., 1996, pp. 132–144.
- [24] C. J. Hawker, E. E. Malmström, C. W. Frank, J. P. Kampf, J. Am. Chem. Soc. 1997, 119, 9903-9904.
- [25] T. H. Mourey, S. R. Turner, M. Rubinstein, J. M. J. Fréchet, C. J. Hawker, K. L. Wooley, *Macromolecules* 1992, 25, 2401–2406.
- [26] K. L. Wooley, C. J. Hawker, J. M. Pochan, J. M. J. Fréchet, Macromolecules 1993, 26, 1514-1519.
- [27] E. M. Harth, S. Hecht, B. Helms, E. E. Malmström, J. M. J. Fréchet, C. J. Hawker, J. Am. Chem. Soc. 2002, 124, 3926–3938.
- [28] S. Abele, K. Vögtli, D. Seebach, Helv. Chim. Acta 1999, 82, 1539-1558.
- [29] S. Abele, P. Seiler, D. Seebach, Helv. Chim. Acta 1999, 82, 1559-1571.

- [30] A. Boeijen, R. M. J. Liskamp, Eur. J. Org. Chem. 1999, 2127 - 2135.
- [31] B. T. Burlingham, T. S. Widlanski, J. Am. Chem. Soc. 2001, 123, 2937-2945.
- [32] A. Cheguillaume, A. Salaün, S. Sinbandhit, M. Potel, P. Gall, M. Baudy-Floch, P. Le Grel, J. Org. Chem. 2001, 66, 4923 - 4929.
- [33] T. D. W. Claridge, J. M. Goodman, A. Moreno, D. Angus, S. F. Barker, C. Taillefumier, M. P. Watterson, G. W. J. Fleet, Tetrahedron Lett. 2001, 42, 4251-4255.
- [34] G. Guichard, V. Semetey, C. Didierjean, A. Aubry, J.-P. Briand, M. Rodriguez, J. Org. Chem. 1999, 64, 8702-8705.
- [35] T. Haack, M. W. Peczuh, X. Salvatella, J. Sánchez-Quesada, J. de Mendoza, A. D. Hamilton, E. Giralt, J. Am. Chem. Soc. **1999**, *121*, 11813–11820.
- [36] H.-S. Lee, F. A. Syud, X. Wang, S. H. Gellman, J. Am. Chem. Soc. 2001, 123, 7721-7722.
- [37] M. C. F. Monnee, M. F. Marijne, A. J. Brouwer, R. M. J. Liskamp, Tetrahedron Lett. 2000, 41, 7991-7995.
- [38] A. T. Lee, A. J. McHugh, Macromolecules 2001, 34, 7127 - 7134.
- [39] T. Glauser, C. M. Stancik, M. Möller, S. Voytek, A. P. Gast, J. L. Hedrick, Macromolecules 2002, 35, 5774-5781.
- [40] M. E. Mackay, Y. Hong, M. Jeong, B. M. Tande, N. J. Wagner, S. Hong, S. P. Gido, R. Vestberg, C. J. Hawker, Macromolecules **2002**, 35, 8391-8399.
- [41] A. Glättli, X. Daura, D. Seebach, W. F. van Gunsteren, J. Am. Chem. Soc. 2002, 124, 12972-12978.

© 2003 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

- [42] D. J. Pochan, L. Pakstis, E. Huang, C. Hawker, R. Vestberg, J. Pople, Macromolecules 2002, 35, 9239-9242.
- [43] S. Hanessian, X. Luo, R. Schaum, S. Michnick, J. Am. Chem. Soc. 1998, 120, 8569-8570.
- [44] G. R. Newkome, J. K. Young, G. R. Baker, R. L. Potter, L. Audoly, D. Cooper, C. D. Weis, K. F. Morris, C. S. Johnson, Jr., Macromolecules 1993, 26, 2394-2396.
- [45] J. K. Young, G. R. Baker, G. R. Newkome, K. F. Morris, C. S. Johnson, Jr., Macromolecules 1994, 27, 3464-3471.
- [46] J. Zhang, J. S. Moore, Z. Xu, R. A. Aguirre, J. Am. Chem. Soc. **1992**, 114, 2273-2274.
- [47] Y. S. Klausner, M. Bodansky, Synthesis 1972, 453-463.
- [48] B. Belleau, G. Malek, J. Am. Chem. Soc. 1968, 90, 1651–1652.
- [49] M. Bodanzsky, A. Bodanzsky, The Practice of Peptide Synthesis, Reactivity and Structure Concepts in Organic Chemistry, Springer Verlag, New York, 1984, p. 145.
- [50] H. A. Bruson, U. S. Pat. 2,401,607, **1946**.
- [51] X. A. Dominguez, I. C. Lopez, R. Franco, J. Org. Chem. 1961, 26, 1625.
- [52] S. J. Allen, J. G. Napler, U. S. Pat. 2,502,548, 1950.
- [53] J. P. Adams, D. S. Box, J. Chem. Soc., Perkin Trans. 1 1999, 749 - 764.
- [54] C. D. Weis, G. R. Newkome, *Synthesis* **1995**, 1053–1065.
- [55] R. M. Silverstein, G. C. Bassler, T. C. Morrill, Spectrometric Identification of Organic Compounds, 5th ed., John Wiley & Sons, Inc., New York, NY, 1991, p. 183.
- [56] G. R. Newkome, X. Lin, Macromolecules 1991, 24, 1443-1444.
- [57] G. R. Newkome, C. D. Weis, Org. Prep. Proced. Int. 1996, 28, 242 - 246.

Received April 15, 2003